

REMARKS

Claims 3, 5-11, 17, 18 and 22-31 presently appear in this case. No claims have been allowed. The official action of July 6, 2005, has now been carefully studied.

Reconsideration and allowance are respectfully urged.

Briefly, the present invention relates to a method for preparing a conjugate of (1) at least one therapeutic agent for joint diseases, which is bonded via a spacer to (2) hyaluronic acid, a hyaluronic acid derivative or a salt thereof (referred to below as "HA"), wherein a carboxyl group of the HA and an amino group of the spacer form an amide bond (claim 11). The invention also relates to a method for treating a patient having a joint disease by administering a pharmaceutical composition containing a pharmaceutically effective amount of the conjugate (claim 17) and a method of treating a joint disease in a patient in need thereof by administering a pharmaceutical composition to the patient in an amount sufficient for such treatment, wherein such pharmaceutical composition includes the conjugate (claim 22).

The conjugate in claims 17 and 22 has a superior effect for the treatment of joint diseases. The conjugate of the present invention can be retained without being dissociated or decomposed at the target site (i.e., a joint cavity) for a long period of time and thus, hyaluronic acid

Appln. No. 09/700,879
Amdt. dated December 6, 2005
Reply to Office action of July 6, 2005

and the therapeutic for joint disease exhibit their own efficacies to produce the desired synergism at the target site with less frequency of administration.

In this respect, the examiner has stated that applicant has supplied no evidence for synergism. However, the synergistic effects of the present invention are sufficiently supported by Figures 5 to 7 of the subject application. Figure 5 shows that the conjugate of the present invention has excellent stability in the bond between HA and the therapeutic agent for joint disease. Figures 6 and 7 show that the conjugate of the present invention has better efficacy and retainability as a therapeutic agent for joint diseases than the therapeutic agent for joint disease and HA that are used either alone or in combination.

Claims 1, 5, 8, 12, 23 and 24 have been rejected under 35 U.S.C. §103(a) as being unpatentable over Akima. Furthermore, claims 1, 3, 5-10, 12, 18, 23 and 24 have been rejected under 35 U.S.C. §103(a) as being unpatentable over Akima and Gallardy.

By the present amendment, the claims relating to the conjugate that were rejected above have been cancelled or converted to method claims. This cancellation and amendment are made without prejudice toward a continuation of the prosecution of conjugate claims at a later date in this or a

Appln. No. 09/700,879
Amdt. dated December 6, 2005
Reply to Office action of July 6, 2005

continuing application. Accordingly, this amendment obviates the above reasons for rejection.

Claims 1, 3, 5-12, 17, 18 and 22-25 have been rejected under 35 U.S.C. §103(a) as being unpatentable over Prestwich, Akima and Gallardy. The examiner states that HA-drug conjugates would have similar utility regardless of the precise physical attachment of the drug to HA, based on the description in Prestwich. This rejection is respectfully traversed.

All of the claims presently appearing in this case are either methods of making or methods of using conjugates having a specific amide bond linkage between the HA and the spacer. All of the conjugates actually prepared in the working examples of Prestwich relate only to those prepared using hydrazides as a spacer. Prestwich requires conjugates prepared by taking advantage of a chemical nature specific to hydrazides. This is clear from the description about the advantages of a pendant hydrazido group on the HA molecule as shown in column 3, line 62, to column 4, line 16, and column 4, line 57, to column 5, line 11. In particular, Prestwich discloses that, because of the low pKa values of the protonated forms of the dihydrazides, over 90% of the species present are unprotonated under the coupling conditions; and the low pKa values of the hydrazido group allows subsequent coupling and crosslinking reactions that can be conducted

Appln. No. 09/700,879
Amdt. dated December 6, 2005
Reply to Office action of July 6, 2005

under conditions of neutral pH that are not detrimental to the molecular weight of the HA molecule. As to the pKa, Prestwich teaches in column 9, lines 38-48, that the preferred dihydrazides have a pKa for the protonated form, less than 8, preferably in the range of 1 to 7 and most preferably in the range of 2 to 6. In contrast to that, the pKa of the protonated amino groups of amino acids is in the range of 12 to 13.

The above disclosures in Prestwich clearly show that hydrazides are distinguished from other substituents, such as an amino group of the spacer used in the present invention. In particular, the coupling reaction attained due to the low pKa of hydrazides teaches away from the coupling reaction in which amino groups, having a higher pKa, are involved.

Therefore, the examiner's statement, "regardless of the precise physical attachment of the drug to HA," is incorrect. The precise physical attachment is, indeed, critical to the process of Prestwich.

In addition, Prestwich discloses that diamines as a spacer do not cause a successful coupling reaction with HA. Specifically, Prestwich discloses in column 7, lines 26-41, that when aliphatic or aromatic diamines were reacted with the HA carboxylic group under coupling reaction conditions, the products formed were only N-acylurea adducts of HA and EDC, citing Pouyani et al, J Am Chem Soc 114:5972-5976 (1992), a

Appln. No. 09/700,879
Amdt. dated December 6, 2005
Reply to Office action of July 6, 2005

copy of which is attached hereto. The failure to prepare the desired conjugate in the case of using diamines as a spacer as disclosed in Prestwich teaches away from the conjugate of the present invention in which a carboxyl group of HA and an amino group of a spacer must form an amide bond.

In view of the above descriptions, Prestwich offers no reasonable expectation for success of the conjugate of the present invention in which a carboxyl group of HA and an amino group of a spacer form an amide bond. Furthermore, the present invention has advantages or unexpected effects over those of Prestwich as explained below.

In the method of producing the HA-drug conjugates disclosed in Prestwich, dihydrazides as a reaction reagent are obtained from NH_2NH_2 (see column 8, lines 46-48). This compound is hydrazine, which is known to be toxic. See, for example, the attached description of "hydrazine" from the Wikipedia on-line encyclopedia.

On the other hand, the method for producing HA-drug conjugates of the present invention in claim 11, which uses amines as a spacer has an advantage over that of Prestwich in that no toxic reagent such as hydrazine is used.

In addition, the only conjugate specifically made in the examples of Prestwich is the hydrazido-HA 1b referred to in Examples 2 (column 26, line 7) and 3 (column 26, line 59) and hydrazido-HA 1b corresponds to (1b) in Example 1 (column

Appln. No. 09/700,879
Amdt. dated December 6, 2005
Reply to Office action of July 6, 2005

24, line 64, to column 65, line 42), which is prepared from HA hexasaccharide (column 24, line 64). This is a low molecular weight HA oligomer prepared according to column 16, line 65, to column 17, line 25. This lower molecular weight HA hexasaccharide disclosed in Prestwich is not suitable as a therapeutic agent for joint disease as the latter requires a certain molecular weight for achieving retainability. HA in the present invention, as a therapeutic agent for joint disease, has an average molecular weight of 100,000 to 10,000,000 as explained in the present specification at page 22, lines 7-9.

Prestwich also discloses in column 10, line 50, that drugs may be covalently attached through the intermediacy of hydrolytically and/or enzymatically labile bonds. This allows the preparation of controlled release formulations.

Hydrazides, amides, etc., are listed as examples of labile bonds. Prestwich by no means suggests retainability of the HA-drug conjugate in which HA and the drug are kept joined to each other via the spacer.

Therefore, Prestwich does not suggest at all that the conjugate disclosed is used for a therapeutic agent for joint disease, still less does it suggest retainability of the HA-drug conjugate in which HA and the drug are kept joined to each other via the spacer without being dissociated or

Appln. No. 09/700,879
Amdt. dated December 6, 2005
Reply to Office action of July 6, 2005

decomposed at the target site (i.e., a joint cavity) for a long period of time.

It cannot be expected from Prestwich, even in combination with Akima and Gallardy, that the conjugate of the present invention has better efficacy and retainability as a therapeutic agent for joint diseases than the therapeutic agent for joint disease or HA used alone or in combination, as shown in Figs. 5-7.

Accordingly, the present invention is by no means obvious from the cited references. Reconsideration and withdrawal of this rejection are therefore respectfully urged.

All of the claims now present in the case fully define over the references of record. Reconsideration and allowance are therefore earnestly solicited.

Respectfully submitted,

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Solid-State NMR of *N*-Acylureas Derived from the Reaction of Hyaluronic Acid with Isotopically-Labeled Carbodiimides

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Abstract: Hyaluronic acid (HA) is a naturally-occurring linear polysaccharide consisting of alternating D-glucuronic acid and *N*-acetyl-D-glucosamine residues. Reaction of the carboxyl group of the glucuronate residues with 1-ethyl-3-(3-(dimethylamino)propyl)carbodiimide (EDC) in the presence of primary amines yielded only the *N*-acylurea adducts rather than the expected amide coupling products. To determine the nature of this linkage unambiguously and to deduce the primary structure of the *N*-acylurea products, ¹³C- and ¹⁵N-labeled 1-ethyl-3-(3-(dimethylamino)propyl)carbodiimide were synthesized. The isotopically-labeled carbodiimides were coupled to the carboxyl group of HA (molecular size ca. 2 000 000 Da) in water at pH = 4.75. The modified polysaccharides were then isolated, purified, and examined by cross polarization and magic angle spinning (CP-MAS) solid-state ¹³C and ¹⁵N NMR. The chemical shifts and states of protonation of the nitrogens confirmed the presence of two isomeric *N*-acylureas in unequal amounts and ruled out the presence of any rearranged *O*-acylurea product.

Introduction

Hyaluronic acid (HA, Figure 1) is a viscoelastic biomaterial that, when present in concentrations as low as 0.1%, can be responsible for 80% of the total viscosity of certain biological fluids. HA is abundant in connective tissue, in the extracellular matrix, and in the vitreous body of the eye.¹ Hyaluronate is also believed to play an important role in cell proliferation and in the control of morphogenesis.² The use of HA esters as a drug delivery system has been reported recently.³ Hyaluronic acid is also an important product in wound-healing⁴ and in cosmetic⁵ preparations.

Chemical modifications of HA have targeted both the hydroxyl and carboxyl functionalities.⁶ However, in most cases, an unambiguous characterization of the final products was not provided.^{7,8} A versatile chemical modification of HA would employ a bifunctional linker, e.g., one in which a variety of therapeutic agents could be attached to a sterically-accessible primary amine after modification of the HA. However, reaction of HA with 1-ethyl-3-(3-(dimethylamino)propyl)carbodiimide (EDC)⁹ in the presence of an excess of either an aliphatic amine or 1,6-diaminohexane failed to provide polymeric materials in which the amine had been incorporated. Instead, the modified HA was tentatively characterized by ¹H NMR as the *N*-acylurea adduct of HA and EDC. On the basis of this observation, we hypothesized that functionalized carbodiimides could be designed, synthesized, and coupled to HA under similar conditions to provide polymeric adducts bearing appropriate functional groups. These functionalized biopolymers could then serve as potentially nonimmunogenic, biodegradable drug delivery agents.

In order to extend the carbodiimide methodology to further modifications of HA, it was necessary (a) to determine the primary structure of the EDC-modified HA unambiguously, (b) to differentiate between the possible isomeric *N*-acylurea or *O*-acylurea products, and (c) to quantify the degree of coupling. Thus, ¹³C- and ¹⁵N-labeled EDC were synthesized, and the HA adducts of these EDC isotopomers were prepared. Herein we report the details of the synthesis of the labeled carbodiimides, the synthesis and purification of the HA-EDC adducts, and the results of the CP-MAS solid-state ¹³C NMR and ¹⁵N NMR studies performed on the isolated, fibrous polymeric adducts.

Solid-state NMR has become a valuable analytical tool for the characterization of insoluble or high molecular weight biomaterials, such as fibrous¹⁰ and membrane¹¹ proteins, native DNA,¹² and insect exoskeletons.¹³ Cross polarization combined with magic angle spinning (CP-MAS)¹⁴ gives well-resolved ¹³C or ¹⁵N NMR spectra, with signal intensities that are in general quantitative or nearly so, and without the extreme solubilization procedures that

would be necessary to obtain solution NMR spectra. While the extensive panoply of techniques available to the solution NMR spectroscopist cannot in general be applied to the solid state, interpretation of chemical shift changes^{11,12} and rotational resonance methods¹⁵ nonetheless can give accurate chemical and structural information. The present work demonstrates the usefulness of these techniques for characterizing the chemical modification of a biopolymer at levels as low as 1% of all monomer units.

Experimental Section

General Procedures. High-resolution ¹H and ¹³C NMR solution spectra were recorded on a General Electric QE-300 300-MHz spectrometer. Chemical shifts are given in ppm using residual CHCl₃ (7.26 ppm) as an internal standard, unless otherwise indicated. IR spectra were determined with a Perkin-Elmer 1600 FTIR instrument. [¹³C]Carbon disulfide and [¹⁵N]ethylamine hydrochloride were purchased from Cambridge Isotope Laboratories. Hyaluronic acid (Amvisc), obtained as its sodium salt (sodium hyaluronate, batch no. 5905) was provided by MedChem Products, Inc. 3-(Dimethylamino)propylamine, thiophosgene, and methyl chloroformate were purchased from Aldrich Chemical Co.

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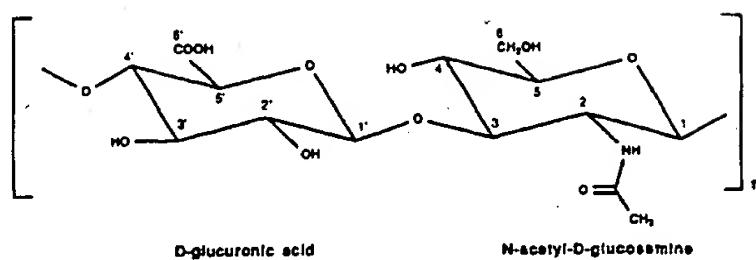


Figure 1. The repeating disaccharide unit in hyaluronic acid.

All solvents used were Optima (HPLC grade), obtained from Fisher Chemical Co.

[¹³C]Ethyl Isothiocyanate (1). In an ice-cooled 2-mL conical vial equipped with a condenser and a stir bar were placed, via syringe, [¹³C]carbon disulfide (500 mg, 6.0 mmol, 99% ¹³C enriched) and 0.59 mL of 11.25 M NaOH (264 mg, 6.0 mmol). To this mixture was added ethylamine (297 mg, 6.0 mmol) as a 33% solution in water. The mixture was heated at 80 °C for 2 h. The resulting red solution was cooled down to 30–40 °C, and methyl chloroformate (620 mg, 6.0 mmol) was added to the mixture via syringe. The ethyl isothiocyanate separated as a brown oil on top of the reaction mixture and was carefully removed by pipet to a small test tube, dried with magnesium sulfate, and subjected to bulb-to-bulb distillation using a Kugelrohr oven with collection in a U-tube at –78 °C to give the [¹³C]ethyl isothiocyanate 1 (180 mg, 31% yield), bp 130–140 °C; ¹H NMR (CDCl₃) δ 1.36 (t, 3 H, J = 7.0 Hz, CH₃), 3.53–3.57 (m, 2 H, CH₂CH₂); ¹³C NMR (CDCl₃) δ 129.4 (N=—¹³C=S), 40.1 (CH₃CH₂), 15.5 (CH₃); IR (neat) 2073 cm^{−1} (N=—¹³C=S).

[2-¹³C]-1-Ethyl-3-(3-(dimethylamino)propyl)thiourea (3). [¹³C]Ethyl isothiocyanate (1) (150 mg, 1.70 mmol) was transferred with 1 mL of chloroform to a 2-mL conical vial. To this mixture was added via syringe an equimolar amount of 3-(dimethylamino)propylamine (2) (173 mg, 1.70 mmol). The mixture was stirred overnight at room temperature under nitrogen. The chloroform was removed by rotary evaporation, and the crude thiourea (300 mg, 96% yield) was used in the next reaction without further purification: ¹H NMR (CDCl₃) δ 1.18 (t, 3 H, J = 7.3 Hz, CH₃CH₂NH¹³C(S)NH), 1.60–1.69 (m, 2 H, CH₂CH₂CH₂N(CH₃)₂), 2.21 (s, 6 H, N(CH₃)₂), 2.32–2.41 (m, 2 H, CH₂CH₂CH₂N(CH₃)₂), 3.30–3.50 (m, 4 H, CH₂CH₂NH¹³CSNHCH₂); ¹³C NMR (CDCl₃) δ 180.5 (NH¹³CSNH).

[2-¹³C]-1-Ethyl-3-(3-(dimethylamino)propyl)carbodiimide (4). The labeled thiourea 3 (280 mg, 1.50 mmol) was dissolved in 2 mL of acetone. To this mixture was added HgO (352 mg, 1.60 mmol), and the mixture was heated at 80 °C for 3 h, gradually turning black upon the formation of HgS. The reaction mixture was filtered through Celite, concentrated by rotary evaporation, redissolved in chloroform, dried (MgSO₄), and transferred to the microdistillation apparatus. The chloroform was evaporated with a gentle stream of nitrogen, and the residue was subjected to bulb-to-bulb distillation under reduced pressure. [2-¹³C]EDC (132 mg) was isolated as a colorless oil in 57% yield after distillation: ¹H NMR (CDCl₃) δ 1.22 (t, 3 H, J = 7.2 Hz, CH₃CH₂N=—¹³C=S), 1.68–1.73 (m, 2 H, CH₂CH₂CH₂N(CH₃)₂), 2.22 (s, 6 H, N(CH₃)₂), 2.34 (t, 2 H, J = 7.1 Hz, CH₂CH₂CH₂N(CH₃)₂), 3.21–3.27 (m, 4 H, CH₂CH₂N=—¹³C=S), ¹³C NMR (CDCl₃) δ 16.6 (CH₃CH₂N=—¹³C=S), 29.3 (=NCH₂CH₂CH₂N(CH₃)₂), 41.4 (CH₃CH₂N=—¹³C=S), 44.8 (=NCH₂CH₂CH₂N(CH₃)₂), 45.4 (N(CH₃)₂), 56.8 (=NCH₂CH₂CH₂N(CH₃)₂), 140.5 (N=—¹³C=S); IR (neat) 2073 cm^{−1} (N=—¹³C=S).

3-(Dimethylamino)propyl Isothiocyanate (5). A mixture of NaHCO₃ (4.9 g, 58 mmol), 3-(*N,N*-dimethylamino)propylamine (1.0 g, 9.7 mmol), 40 mL of H₂O, and 50 mL of HPLC grade chloroform was stirred for 10 min. Thiophosgene (1.7 g, 14.5 mmol) was added to this mixture via syringe, and the reaction mixture was stirred for 1 h at room temperature, turning gradually from red to yellow. The chloroform layer was dried (MgSO₄), filtered, and concentrated by rotary evaporation. The desired product 5 was obtained as an orange oil in 35% yield. The crude product was used without further purification: ¹H NMR (CDCl₃) δ 1.83 (m, 2 H, (CH₃)₂NCH₂CH₂CH₂), 2.21 (s, 6 H, N(CH₃)₂), 2.37 (t, 2 H, J = 6.8 Hz, (CH₃)₂NCH₂CH₂CH₂), 3.58 (t, 2 H, J = 6.6 Hz, S=C=NCH₂CH₂CH₂); IR (neat) 2108 cm^{−1} (N=C=S).

[1-¹⁵N]-1-Ethyl-3-(3-(dimethylamino)propyl)thiourea (6). Crude isothiocyanate 5 (450 mg, 3.10 mmol) was dissolved in 4 mL of chloroform. To this mixture were added [¹⁵N]ethylamine hydrochloride (250 mg, 3.10 mmol, 99% ¹⁵N enriched) and an equimolar amount of triethylamine (330 mg, 3.10 mmol). The ethylamine hydrochloride was initially only partially soluble in the system; however, after stirring overnight at room temperature, no traces of solid were observed, indicating that it had gradually dissolved. The reaction mixture was washed with 5% NaOH, extracted with chloroform, dried (MgSO₄), and concentrated by rotary evaporation, yielding 610 mg (quantitative) of crude thiourea 6, which

was used without further purification: ¹H NMR (CDCl₃) δ 1.17–1.20 (m, 3 H, CH₃CH₂¹⁵NCSNH), 1.69 (m, 2 H, CH₂CH₂CH₂N(CH₃)₂), 2.23 (s, 6 H, CH₂CH₂CH₂N(CH₃)₂), 2.41 (m, 2 H, CH₂CH₂CH₂N(CH₃)₂), 3.20–3.50 (m, 4 H, CH₃CH₂¹⁵NCSNHCH₂).

[1-¹⁵N]-1-Ethyl-3-(3-(dimethylamino)propyl)carbodiimide (7). The same procedure described above for the synthesis of [¹³C]EDC (4) was employed. Thus, thiourea 6 (600 mg, 3.20 mmol) was subjected to dehydrosulfurization with HgO (750 mg, 3.50 mmol) in refluxing acetone to afford 210 mg of [¹⁵N]EDC (7) in 43% yield after distillation: ¹H NMR (CDCl₃) δ 1.20–1.24 (m, 3 H, CH₃CH₂¹⁵N=C=N), 1.68–1.73 (m, 2 H, CH₂CH₂CH₂N(CH₃)₂), 2.20 (s, 6 H, N(CH₃)₂), 2.29–2.34 (t, 2 H, J = 7.5 Hz, CH₂CH₂CH₂N(CH₃)₂), 3.18–3.25 (m, 4 H, CH₃CH₂¹⁵N=C=NCH₂); ¹³C NMR (CDCl₃) δ 16.6, 29.3, 41.4, 44.8, 45.5, 56.9; IR (neat) 2108 cm^{−1} (N=—¹⁵N=C=N).

General Procedure for Coupling of Labeled EDC to Sodium Hyaluronate. Sodium hyaluronate was dissolved in water such that the concentration of the HA solution was approximately 4 mg/mL. The pH of the solution was adjusted to 4.75 using 0.01 N HCl. A 0.5 mg/mL solution of the labeled carbodiimide (either 4 or 7) in H₂O was prepared, and the pH was adjusted to 5.00 by addition of 0.1 N HCl. The carbodiimide solution was then added dropwise to the HA solution over a period of 15 min. The pH of the reaction mixture was maintained at 4.75 throughout the entire course of the reaction by titration with 0.01 N HCl. The proton consumption was measured in order to quantitate the degree of coupling. For workup, sodium chloride (5% w/v) was added to salt out the modified HA. The mixture was then added to 3 volumes of 95% ethanol, and the modified HA was obtained as a stringy white precipitate. The precipitate was squeezed dry and redissolved in the original volume of water for a second precipitation. The precipitation and redissolution sequence was repeated three times to ensure the removal of all low molecular weight organic impurities (unreacted starting material). The resulting solution of HA was clear and viscous and was placed on a lyophilizer for 48 h.

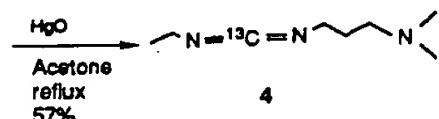
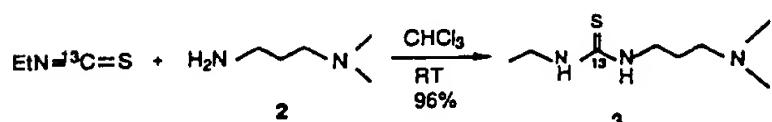
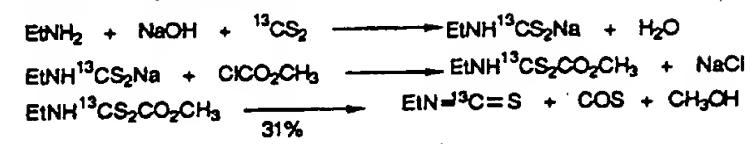
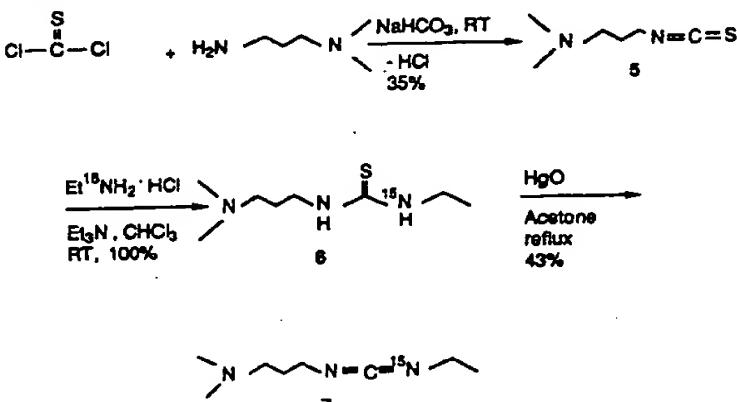
The ¹H NMR samples were prepared by dissolving 10 mg of modified HA in 1 mL of D₂O and adding 10 μ L of 1 N NaOD to decrease the viscosity of the solution. The standard used was 3-(trimethylsilyl)propanesulfonic acid, sodium salt hydrate, set at δ 0.00 ppm. The reactions were carried out at room temperature with carbodiimides 4 and 7 and also at 4 °C with compound 7. The maximum theoretical coupling was 20% based on molar equivalents used. However, 3.0 and 4.8% coupling were observed for the reactions of 4 and 7 with HA, respectively, based on H⁺ consumption data. Proton consumption data was not available for the reaction of 7 with HA at 4 °C. The reaction times were 2 h at room temperature and 16 h at 4 °C; longer reaction times did not give increased coupling: ¹H NMR (D₂O) δ 1.09 (t, 3 H, CH₃CH₂N), 1.65 (m, 2 H, NCH₂CH₂CH₂N(CH₃)₂), 2.19 (s, 6 H, N(CH₃)₂), 2.34 (t, 2 H, NCH₂CH₂CH₂N(CH₃)₂), 3.12 (m, 4 H, CH₃CH₂NCONHCH₂).

Solid-State NMR Spectroscopy. Solid-state NMR spectra were obtained using a home-built spectrometer operating at a field of 7.1 T (301.42-MHz ¹H frequency). Both ¹³C and ¹⁵N spectra were acquired with cross polarization and magic angle spinning¹⁴ using a home-built probe incorporating Doty Scientific stator and rotors. Typical sample quantities were 250 mg. Spinning speeds were 3–4.5 kHz. Proton, ¹³C, and ¹⁵N rotating frame frequencies were 55–60 kHz during both cross polarization and decoupling. Recycle delays were usually 3 s, which was quite adequate to allow full relaxation of the proton spin system, and were limited by sample heating rather than by relaxation. Usually, 10 000–30 000 transients were averaged/spectrum. The cross polarization contact time was 1 ms for ¹³C; a contact time of 3 ms was used for ¹⁵N to ensure full magnetization transfer even to unprotonated nitrogen species. Under these experimental conditions, the sample temperature was approximately 27 °C. Protonated and unprotonated resonances were distinguished using the delayed decoupling experiment;¹⁶ delays for dipolar dephasing were 40 μ s for ¹³C and 80 μ s for ¹⁵N; in both cases these timings were found empirically to create a null in the signal of immobile singly-protonated species. Other assignments were made by comparison of solid- and solution-state chemical shifts, or by comparing the spectra of labeled modified hyaluronate polymers with unlabeled modified samples on the one hand, and with unmodified hyaluronate on the other. ¹³C chemical shifts were referenced to the downfield line of adamantane and converted to the tetramethylsilane scale by adding 38.56 ppm.¹⁷ ¹⁵N chemical shifts were likewise measured with respect to solid NH₄Cl, and 14.8 ppm was added to express these shifts with respect to 5.6 M NH₄Cl in H₂O.¹⁸

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Scheme I. Synthesis of [2-¹³C]EDC**Scheme II. Synthesis of [1-¹⁵N]EDC****Results and Discussion**

[2-¹³C]EDC selectively labeled at the central sp-hybridized carbon was synthesized in three steps (Scheme I). The reaction of [¹³C]CS₂ with ethylamine and sodium hydroxide, followed by treatment with methyl chloroformate, afforded [¹³C]ethyl isothiocyanate (1) in 31% yield after bulb-to-bulb distillation.¹⁹ Reaction of [¹³C]ethyl isothiocyanate with 3-(dimethylamino)-propylamine (2) provided thiourea 3 in 96% yield. Thiourea 3 was subjected to dehydrosulfurization²⁰ to give the [2-¹³C]-labeled carbodiimide 4 in 32% yield after bulb-to-bulb distillation.

The synthesis of [1-¹⁵N]EDC labeled on the N-ethyl nitrogen of the carbodiimide was accomplished in three steps (Scheme II). Treatment of thiophosgene with 3-(dimethylamino)propylamine in the presence of sodium bicarbonate afforded 3-(dimethylamino)propyl isothiocyanate (5) in 35% yield.²¹ Treatment of isothiocyanate 5 with [¹⁵N]ethylamine hydrochloride in the presence of an equimolar amount of triethylamine afforded thiourea 6 in quantitative yield. Dehydrosulfurization of thiourea 6 with HgO in refluxing acetone afforded [1-¹⁵N]EDC (7) in 43% yield after bulb-to-bulb distillation.

The isotopically-labeled carbodiimides were each coupled to HA at pH 4.75 at room temperature.²² The pH of each reaction was maintained at 4.75 by addition of 0.01 N HCl. This pH was chosen, since under these conditions a sufficient number of free carboxylates are present, while at the same time the solution is sufficiently acidic to promote carbodiimide activation. At lower pH, HA solubility and integrity were compromised; at higher pH, insufficient reaction with EDC was observed. Subsequent studies with HA fragments of defined length (2–5 disaccharide units)

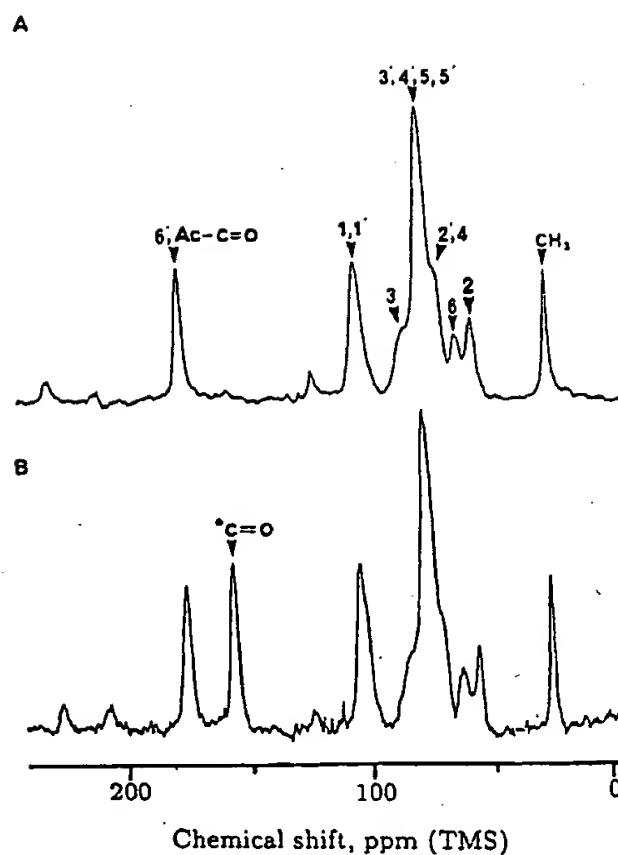


Figure 2. (A) ¹³C CP-MAS NMR spectrum of native hyaluronate. (B) ¹³C CP-MAS NMR spectrum of [¹³C]EDC-HA adduct.

Table I. Chemical Shifts, Line Widths at Half Height, Relative Intensities (Calculated Intensities in Parentheses), and Assignment of the ¹³C Spectrum of Native Hyaluronic Acid^a

chemical shift (ppm)	line width (ppm)	relative intensity	assignment
24.7	1.9	0.87 (1)	acetyl-CH ₃
55.7	3.8	1.00 (1)	2
62.3	4.9	0.96 (1)	6
70.1	4.5	1.59 (1-2)	2', 4
76.1	4.8	4.52 (4-5)	3', 4', 5, 5'
84.2	4.8	1.05 (1)	3
102.7	5.3	2.09 (2)	1, 1'
175.2	3.6	1.92 (2)	6', acetyl-CO

^aChemical shifts are referenced to external tetramethylsilane (TMS) and are uncorrected for bulk susceptibility.

have confirmed this pH as that which provides the opportunity for optimum coupling.²³ The maximum theoretical coupling was 20% based on the molar equivalents used. This value was selected to obtain sufficient coupling for observation of the ¹⁵N and ¹³C resonances for the EDC-HA adduct balanced against the declining efficiency of coupling at higher percentages of modification.

The reaction of a carbodiimide with a carboxyl group generally proceeds through the addition of the free carboxylate to one of the double bonds of the diimide system to give an O-acylurea product.²⁴ In the presence of a nucleophile the acyl-nucleophile product will form, plus the urea of the carbodiimide. The second pathway, which accounts for hindered O-acylureas and occurs in the absence of added nucleophiles, is for the O-acylurea to rearrange to the more stable N-acylurea through an intramolecular acyl transfer. The scope of this chemical modification of high molecular weight HA by a variety of carbodiimides has been described recently.²⁵

The ¹³C CP-MAS spectrum of native hyaluronate is shown in Figure 2A. It was obtained at a spinning frequency of 3.63 kHz.

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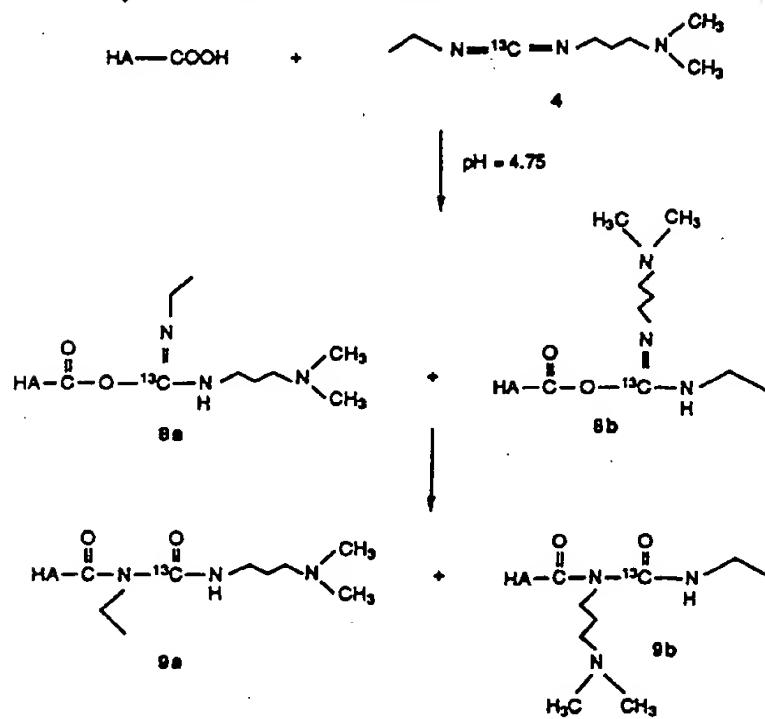
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Scheme III. Possible Reaction Pathways for Coupling of [2-¹³C]EDC with HA, Showing Regioisomeric *O*-Acylurea Adducts and *N*-Acylurea Rearrangement Products



Under these conditions at a field strength of 7.1 T, only the carbonyl groups of hyaluronate have significant rotational sideband intensity; these sidebands are labeled. The remaining lines were fit to Lorentzians, using a deconvolution program to disentangle the overlapped resonances in the main sugar carbon region. The chemical shifts, line widths, and relative intensities thus obtained are given in Table I. The summed intensity of all peaks, including the carbonyl sidebands, was normalized to 14 carbons. The unprotonated and methyl carbons were identified using a delay without decoupling sequence,¹⁷ with a dipolar dephasing time of 40 μ s. Methylene carbons can also be distinguished from methine carbons by this method if a somewhat shorter dephasing period is used. The remaining carbon signals were assigned by comparison with solution NMR.²⁶

The data in Table I show that CP-MAS intensities obtained for this material are nearly quantitative; all peak intensities fall near integer numbers, with the exception of the strongly overlapped signals at 70.1 and 76.1 ppm. The sum of the latter pair of intensities corresponds nonetheless to that expected for six carbons. We can therefore anticipate that incorporation of isotope levels can be estimated from such spectra with an accuracy of around $\pm 10\%$. The chemical shifts of this fibrous hyaluronate sample are uniformly nearly identical to those observed in solution NMR, with no more than 1 ppm deviation for the clearly resolved resonances or 3 ppm for the overlapped ones. Given that chemical shielding of sugar carbons is highly sensitive to sugar pucker changes or other conformational alterations, with shifts of 10 ppm or greater being noted in some cases,²⁷ these data strongly suggest that the conformations of hyaluronate in solution and in the fibrous state are very similar.

Reaction of hyaluronate with EDC selectively ¹³C-labeled on the carbodiimide residue gave a polymeric adduct (Scheme III). This adduct was precipitated several times from ethanol to remove all traces of unreacted carbodiimide and small molecule byproducts. The purified polymer was studied by ¹³C solid-state NMR, using CP-MAS. In addition to the native HA resonances, a new peak was identified at 156 ppm, consistent with a substituted urea (Figure 2B). The peak intensity corresponds to 0.025 ± 0.002 mol of linker/mol of monomer units. Unfortunately, this chemical

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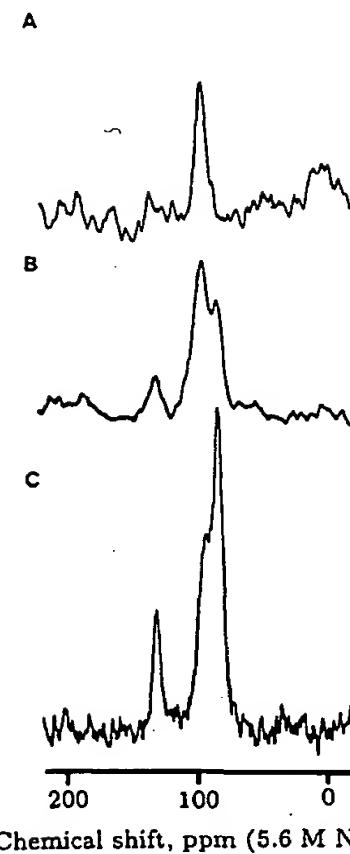
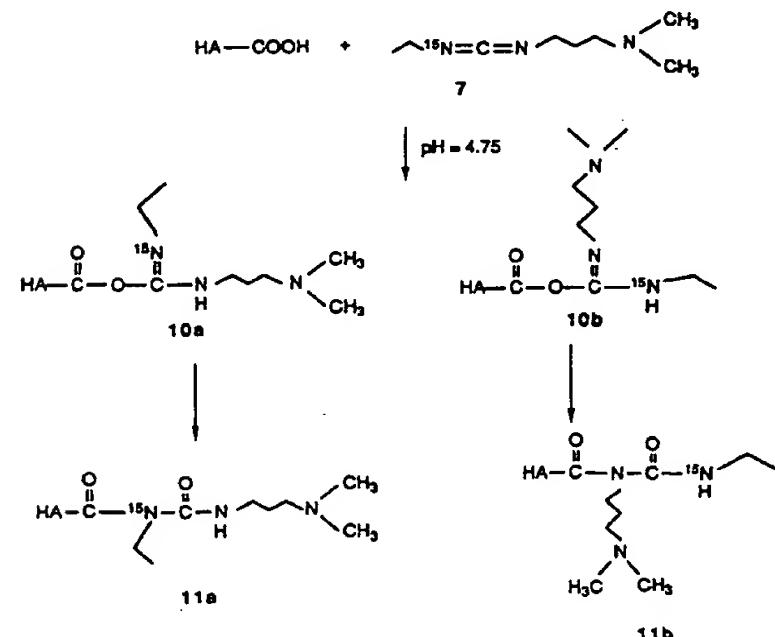


Figure 3. (A) ¹⁵N CP-MAS NMR spectrum of native hyaluronate. (B) ¹⁵N CP-MAS NMR spectrum of [¹⁵N]EDC-HA adduct, 4 °C. (C) ¹⁵N CP-MAS NMR spectrum of [¹⁵N]EDC-HA adduct, 25 °C.

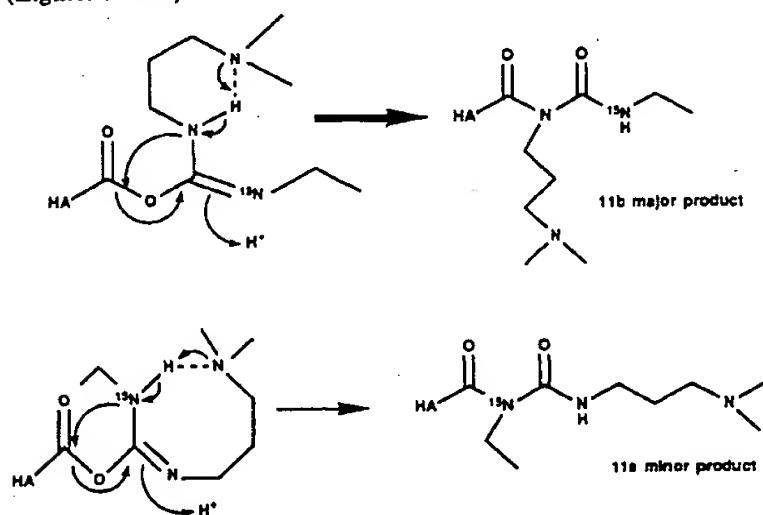
Scheme IV. Possible Reaction Pathways for Coupling of [1-¹⁵N]EDC with HA, Showing Regioisomeric *O*-Acylurea Adducts and *N*-Acylurea Rearrangement Products



shift does not unambiguously discriminate between the two *O*-acylurea isomers 8a and 8b or between them and the two *N*-acylurea isomers 9a and 9b which might be formed from them by rearrangement.

Next, EDC selectively ¹⁵N-labeled on the ethylamine moiety was prepared and coupled with HA (Scheme IV). The ¹⁵N solid-state CP-MAS NMR spectrum now showed three resonances (Figure 3C). The resonance at 96 ppm (relative to 5.6 M NH₄Cl in H₂O) was present also in the native polymer and was therefore assigned to natural abundance ¹⁵N in the acetamido group of HA. Interrupted decoupling experiments showed the high-frequency peak at 133 ppm to be an unprotonated (tertiary) nitrogen. No significant MAS sidebands were observed even at slow spinning

Scheme V. Proposed Route Leading to the Observed Major *N*-Acylurea Product (Heavy Arrow) and to the Minor Product (Lighter Arrow)



speeds, suggesting a small chemical shielding anisotropy (CSA). This tends to rule out formally double-bonded species, such as *O*-acylureas, which usually have large CSAs.¹¹ Additionally, the limited reference data available for *O*-acylureas suggest that they should appear at even higher frequencies. On the other hand, the chemical shift and CSA are entirely consistent with an *N*-acyl-*N*-alkylurea nitrogen such as that in compound 11a, which would be formed by transfer of the acyl group to the ethylamine nitrogen.

The low-frequency peak, at 83 ppm, is a protonated nitrogen species with a chemical shift intermediate between that of a secondary amide and that expected for an alkylurea. Nonetheless, this resonance can be assigned to the latter species, 11b. Its uncharacteristically high chemical shift can be attributed to the transfer of the acyl group to the other available nitrogen, resulting in increased double-bond character of the C-N bond and thus a paramagnetic shift. Deconvolution of the three peaks suggests a total isotope incorporation of 0.020 ± 0.004 mol of isotope/mol of monomer unit, close to that observed with the carbon label.

The effects of a lower reaction temperature (4 °C) on product distribution and on the rate of the rearrangement of *O*-acylurea were examined. Thus, [¹⁵N]EDC was coupled to HA at 4 °C as previously described, and the isolated polymeric adducts were subjected to ¹⁵N solid-state CP-MAS NMR spectroscopy. A longer reaction time (16 h) was allowed to compensate for the reduced reaction rate due to increased viscosity of the HA mixture at low temperature. The relative intensity of the two labeled species was unaffected by this reduction in reaction temperature; furthermore, no additional signals were observed (Figure 3B).

A rationale for the production of unequal amounts of isomeric *N*-acylureas in the coupling reactions of isotopically-labeled EDC is provided in Scheme V. It is likely that the rearrangement of

the *O*-acylurea to an *N*-acylurea is catalyzed intramolecularly by one of the basic amine nitrogens. On the basis of this proposition, the rearrangement that involves a six-membered ring for proton abstraction by the terminal amine should be favored over the alternative eight-membered ring. This model accounts for the observation of 11b as the major *N*-acylated product.

Formation of *N*-acylurea products from the reaction of a carboxylate with a carbodiimide in the absence of a nucleophile is well-documented.²⁸ Modifications of proteins with EDC have been reported to yield *N*-acylurea products that alter the DNA-binding properties of the modified proteins.²⁹ The reactions of *N*-acetylchondrosine, *N*-acetylchondrosine 6-sulfate, and heparin with EDC have been reported,³⁰ in which putative *O*-acylurea products were isolated and which rearranged to the corresponding *N*-acylureas upon alkaline treatment with NaHCO₃.

On the basis of the NMR results, therefore, we conclude that the initial *O*-acylated product of the coupling reaction between EDC and HA rearranges to give a mixture of two *N*-acylated isomers. These two isomers can readily be identified and characterized by solid-state NMR spectroscopy. The present results demonstrate the success of applying CP-MAS toward the detection of low-level chemical modifications in a biopolymer and in assigning unambiguous structures to complex macromolecules. The described approach could potentially be applied to the structural analysis of a variety of soluble and insoluble modified polymeric species. In addition, we have presented the first example of an unambiguous structural characterization of modified hyaluronate using solid-state NMR techniques. We are currently in the process of exploiting the carbodiimide methodology³¹ for further functionalizations of hyaluronic acid, for potential use as a drug delivery vehicle, and for the production of novel biomaterials.

Acknowledgment. We thank MedChem Products, Inc. and the Center for Biotechnology/New York State Foundation for Science and Technology for financial support of this research. We are also grateful to MedChem Product, Inc., for providing sodium hyaluronate, and to D. A. Swann (MedChem) for stimulating our interest in this project. G.S.H thanks the National Institutes of Health (GM39071) for financial support.

Registry No. 1, 141727-00-6; 2, 109-55-7; 3, 141727-01-7; 4, 141727-02-8; 5, 27421-70-1; 6, 141727-03-9; 7, 141727-04-0; Et¹⁵NH₂·HCl, 84051-02-5; ¹³CS₂, 30860-31-2; CICO₂CH₃, 79-22-1; thiophosgene, 463-71-8; hyaluronic acid, 9004-61-9.

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Hydrazine

From Wikipedia, the free encyclopedia.

Hydrazine is a chemical compound with formula N_2H_4 used as a rocket fuel.

Hydrazine is a liquid with weak basic properties similar to ammonia. Due to the alpha effect the nucleophilicity is much stronger than that of ammonia, which makes it more reactive. It can be made by oxidizing ammonia with sodium hypochlorite (the Raschig process). It is a monopropellant rocket fuel.

Hydrazine derivatives 1,1-dimethylhydrazine and 1,2-dimethylhydrazine, in which two of the hydrogen atoms are substituted with methyl groups, are also described as **hydrazines**.

1,1-Dimethylhydrazine is used to make hypergolic (self-igniting) bipropellant rocket fuels.

Health effects

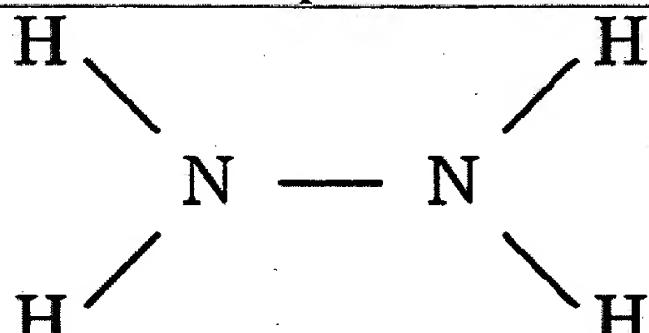
Breathing hydrazines may cause coughing and irritation of the throat and lungs, tremors, or seizures. Breathing hydrazines for long periods may cause liver and kidney damage, as well as serious effects on reproductive organs.

Eating or drinking small amounts of hydrazines may cause nausea, vomiting, uncontrolled shaking, inflammation of the nerves, drowsiness, or coma. Hydrazine is found in chewing tobacco and cigarettes.

Tumors have been seen in many organs of animals that were exposed to hydrazines by ingestion or breathing, but most tumors have been found in the lungs, blood vessels, or colon. 1,2-Dimethylhydrazine has caused colon cancer in laboratory animals following a single exposure.

The Department of Health and Human Services (DHHS) has determined that

Properties



The structure formula of Hydrazine.

General

Name	Hydrazine
Chemical formula	N_2H_4
Appearance	Colourless liquid

Physical

Formula weight	32.0 amu
Melting point	274 K (1 °C)
Boiling point	387 K (114 °C)
Density	1.01 g/ml
Solubility	very soluble

Thermochemistry

$\Delta_f H^0_{\text{gas}}$	95.35 kJ/mol
$\Delta_f H^0_{\text{liquid}}$	50.63 kJ/mol
$\Delta_f H^0_{\text{solid}}$	37.63 kJ/mol
$S^0_{\text{gas}, 1 \text{ bar}}$	238.66 J/mol·K
$S^0_{\text{liquid}, 1 \text{ bar}}$	121.52 J/mol·K
S^0_{solid}	? J/mol·K

Safety

hydrazine and 1,1-dimethylhydrazine are known carcinogens. The International Agency for Research on Cancer (IARC) has determined that hydrazine, 1,1-dimethylhydrazine, and 1,2-dimethylhydrazine are possible human carcinogens. The Environmental Protection Agency (EPA) has determined that hydrazine, 1,1-dimethylhydrazine, and 1,2-dimethylhydrazine are probable human carcinogens.

The American Conference of Governmental Industrial Hygienists (ACGIH) currently lists hydrazine and 1,1-dimethylhydrazine as suspected carcinogens, but has recently recommended that the listing of hydrazine be changed to that of animal carcinogen, not likely to cause cancer to people under normal exposure conditions.

The False Morel contains the chemical gyromitrin, which is metabolized into monomethyl hydrazine inside the body. Consequently, the toxic effects of this mushroom are the same as with hydrazine poisoning.

Use

Hydrazine is used primarily as a chemical intermediate to produce agricultural chemicals, spandex fibers, and antioxidants. Hydrazine is also used as rocket fuel, an oxygen scavenger (corrosion inhibitor) in water boilers and heating systems, a polymerization catalyst, a blowing agent, and as a scavenger for gases. Additionally, it is used for plating metals on glass and plastics and in fuel cells, solder fluxes, and photographic developers. Hydrazine is used as a reactant in fuel cells in the military, as a reducing agent in electrodeless nickel plating, as a chain extender in urethane polymerizations, as a reducing agent in plutonium extraction from reactor waste, and as a water treatment chemical. Hydrazine is also used as a chemical intermediate for blowing agents, photography chemicals, pharmaceuticals, antituberculants, textile dyes, heat stabilizers, explosives, and to make hydrazine sulfate. In addition, it has recently been determined that hydrazine increases the speed of the thin-film transistors used in liquid crystal displays, a discovery that promises to revolutionize the manufacture of LCD computer monitors. Hydrazine in a 70% solution is used to power the EPU (Emergency Power Unit) on the F-16 fighter plane. Hydrazine is also used as low-power propellant for Space Shuttle maneuvers in orbit, as hydrazine can be decomposed from a liquid into gaseous components despite the absence of oxygen, allowing the high pressure gaseous products to be expanded out of a nozzle.

Retrieved from "<http://en.wikipedia.org/wiki/Hydrazine>"

Categories: Bases | Rocket fuels | Nitrogen compounds

Ingestion	Extremely Toxic, possibly carcinogenic
Inhalation	Very dangerous—extremely destructive to the upper respiratory tract
Skin	Can cause severe burns, can be absorbed into bloodstream
Eyes	Can cause permanent damage
More info	Hazardous Chemical Database (http://www.hhmi.org/research/labsafe/lcss/lcss46.html)
LD ₅₀	as low as 25mg/kg

SI units were used where possible. Unless otherwise stated, standard conditions were used.

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